Method 2

Appendix of Real-time quantitative PCR training course.

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This document explains how you can interpret the Ct values from $\Delta\Delta$ Ct method and how you can determine the $\Delta\Delta$ Ct method is applicable for your experiment instead of the conventional calibration curve method.

If you are not sure which method to use, use calibration curve method because it is more reliable for most of the real-time PCR applications.

Experimental condition

- Sample subject: Control (Cont) and Experimental (Exp)
- Target gene: Tar
- Reference gene: *Ref*

Results

- Control sample Ct for Tar (ContCt Tar) was 22, Ct for Ref (ContCt Ref) was 19.
- Experimental sample Ct for Tar (ExpCt Tar) was 25, Ct for Ref (ExpCt Ref) was 18.

$$\Delta \text{ExpCt} = (2^{\text{ExpCt}^{Tar}} / 2^{\text{ExpCt}^{Ref}})$$

= $(2^{25} / 2^{18}) = (2^{25-18}) = 2^{7}$

This calculation assumes that the DNA product becomes 2-fold after each PCR cycle (means 100% PCR efficiency).

This step normalizes the amount of *Tar* by the amount of *Ref* in Exp. (In other wards, *Tar* was 2⁷-times more than *Ref* in Exp).

$$\Delta$$
ContCt = (2^ContCt^{Tar} /2^ContCt^{Ref})
= (2²²/ 2¹⁹) = (2²²⁻¹⁹) = 2³

In the same way, the expression difference is $2^3 = 8$ -fold. This is also assuming 100% PCR efficiency.

This step normalizes the amount of Tar by the amount of Ref in Cont. (Tar was 2^3 -times more than Ref in Cont)

$$\Delta\Delta$$
Ct = (Δ ExpCt/ Δ ContCt) = (2^7 / 2^3) = 2^{7-3} = 2^4

This means *Tar* in Exp requires 4 more rounds of PCR cycle to reach the same amount of *Tar* in Cont.

Therefore Exp would have had $2^4 = 16$ -fold smaller amount of *Tar* compared to that in Cont.

When the amount of *Tar* in Cont is set as 1.0, the amount of *Tar* in Exp is 0.0625 (= 1/16).

Attention!!:

This calculation is applicable only when the PCR efficiency of both *Tar* and *Ref* is 100% (DNA product becomes 2-fold after each PCR cycle).

If the PCR efficiency is less than 100%, you have to change the calculations.

Hypothetical results:

The efficiency was 95% for *Tar* and 93% for *Ref*, meaning that DNA becomes 1.95-fold for *Tar* and 1.93-fold for *Ref* after each PCR cycle.

The calculations change as follows:

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\DeltaExpCt = (1.95^ExpCt<sup>Tar</sup> / 1.93^ExpCt<sup>Ref</sup>) = (1.95<sup>25</sup> / 1.93<sup>18</sup>)

\DeltaContCt = (1.95^ContCt<sup>Tar</sup> / 1.93^ContCt<sup>Ref</sup>) = (1.95<sup>22</sup> / 1.93<sup>19</sup>)

\DeltaCt = (\DeltaExpCt /\DeltaContCt) = (1.95<sup>25</sup> / 1.93<sup>18</sup>) / (1.95<sup>22</sup> / 1.93<sup>19</sup>)

= (1.95<sup>25-22</sup> / 1.93<sup>18-19</sup>) = (1.95<sup>3</sup> / 1.93<sup>-1</sup>)

\rightleftharpoons (7.41 / 0.52) \rightleftharpoons 14.31
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This means that the amount of Tar in Exp was 14.31-fold less than that in Cont so that if the amount of Tar in Cont is set as 1.0, the amount of Tar in Exp is approximately 0.070 (= 1/14.31).

Important rules:

You can use the $\Delta\Delta$ Ct method only when the PCR efficiency of *Tar* and *Ref* is steady, high and almost equal.

How to evaluate the PCR efficiency?

- When you make calibration curves for *Tar* and *Ref*, you will have values of the steadiness (R²), the amplification efficiency and the slope for each calibration curve (refer to the next page).
- If the efficiency is 90% or higher and R² is 0.95 or higher (0.99 or higher is preferred) for both *Tar* and *Ref* and at the same time the difference of slope values between *Tar* and *Ref* is less than 0.1 (means 0.999 is OK but 0.100 is not OK), the experimental condition is good enough to use the ΔΔCt method, otherwise you have to use the calibration curve method.

